KRIBIOLISA™ Darbepoetin (ARANESP) ELISA

REF : KBI1030

Ver 2.2

RUO

Enzyme Immunoassay for the Quantitative Determination of Darbepoetin in human serum and plasma

RUO	For Research Use	REF	Catalog Number
	Store At	LOT	Batch Code
	Manufactured By		Biologi isk
	Expiry D		Consu rating Instructions

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REF KBI1030 Σ 96 tests

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Introduction:

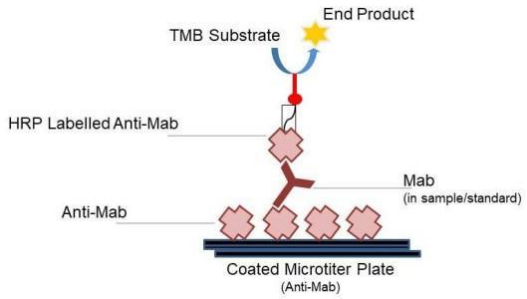
Erythropoietin (EPO) is a heavily glycosylated protein with a molecular weight of about 30,000 - 34,000 Daltons. Human EPO is a polypeptide consisting of 165 amino acids, containing one O-linked and three N-linked carbohydrate chains. The recombinant EPO is a good substitute for the native protein for use in an immunoassay. Darbepoetin is a re-engineered form of erythropoietin, contains 5 N-linked oligosaccharide chains and has a molecular weight of 37,100 Daltons and a carbohydrate composition of 51%.It has a 3-fold longer serum half-life compared to epoetin alpha and epoetin beta.

Intended Use:

The KRIBIOLISA™ Darbepoetin (ARANESP) ELISA is used as an analytical tool for quantitative determination of Darbepoetin in human serum and plasma.

Principle:

The method employs the quantitative sandwich enzyme immunoassay technique Standards or samples are pipetted into microwells pre-coated with Anti-Darbepoetin antibody and Darbepoetin present in the sample and standards are bound by Anti-Darbepoetin antibody. In the second step, Detection Antibody is added and incubated. In the third step, a HRP conjugate is pipetted and incubated. Free HRP conjugate will be removed by washing. Addition of TMB substrate will develop blue color and intensity of blue colour in wells is proportional to the concentration of Darbepoetin present in standard or sample. Color development is then stopped by addition of stop solution. Absorbance is measured at 450 nm.



Materials Provided:

Part	Description	Qty
Anti-Darbepoetin Coated Microtiter Plate	96 well polystyrene microplate (12 strips of 8 wells) coated with Anti-Darbepoetin antibody.	1 x 96 wells
Darbepoetin Standard	Recombinant Darbepoetin standard (Concentration - 1 ug/ml; lyophilized)	2 vials
Darbepoetin detection Antibody	Biotin labelled Darbepoetin detection antibody (Concentration - 3 ug/ml; lyophilized)	1 vial
Streptavidin HRP	Streptavidin HRP conjugate (30ul per vial)	1 vial
(1X) Assay Diluent	Buffered protein base with protein stabilizer and preservatives 0.02% methylisothiazolone and 0.02% bromonitrodioxane.	30 ml
(1X) Sample Diluent	Buffered protein base with protein stabilizer and preservatives 0.02% methylisothiazolone and 0.02% bromonitrodioxane.	50 ml
(1X) Standard Diluent	Buffered protein base with 1:100 dilution human serum with protein stabilizer and preservatives 0.02% methylisothiazolone and 0.02% bromonitrodioxane.	10 ml
(20X) Wash Buffer	20-fold concentrated solution of buffered surfactant with preservative thiomersol < 0.01%. May turn yellow over time.	25 ml
TMB Substrate	Stabilized Chromogen	12 ml
Stop Solution	0.73M Phosphoric Acid	12 ml
Instruction Manual		1 no

Materials to be provided by the End-User:

1. Microtiter Plate Reader able to measure absorbance at 450 nm.
2. Adjustable pipettes and multichannel pipettor to measure volumes ranging from 25µl to 1000µl
3. Deionized (DI) water
4. Wash bottle or automated microplate washer
5. Graph paper or software for data analysis
6. Timer
7. Absorbent Paper

Handling/Storage:

1. All reagents should be stored at 2ºC to 8ºC for stability.
2. All the reagents and wash solutions should be used within 12 months from manufacturing date.
3. Before using, bring all components to room temperature (18-25ºC). Upon assay completion ensure all components of the kit are returned to appropriate storage conditions.
4. The Substrate is light-sensitive and should be protected from direct sunlight or UV sources.

Health Hazard Warnings:

1. Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin.
2. For Research Use Only.



Sample Preparation and Storage:

Blood is taken by venipuncture. Serum is separated after clotting by centrifugation. Plasma can be used, too. Lipaemic, hemolytic or contaminated samples should not be run. Repeated freezing and thawing should be avoided. If samples are to be used for several assays, initially aliquot samples and keep at - 20°C.

Preparation Before Use:
Allow samples to reach room temperature prior to assay. Take care to agitate patient samples gently in order to ensure homogeneity.

Test Sample preparation - Samples have to be diluted 1:100 (v/v), e.g. 1 ul sample + 99 ul sample diluent prior to assay. The samples may be kept at 2 - 8°C for up to three days. Long-term storage requires -20°C.

Reagent Preparation (all reagents should be diluted immediately prior to use):

1. Label any aliquots made with the kit Lot No and Expiration date and store it at appropriate conditions mentioned.
2. Bring all reagents to Room temperature before use.
3. To make Wash Buffer (1X); dilute 25 ml of (20X) Wash Buffer in 475 ml of DI water.
4. **Standards Preparation**: Reconstitute the lyophilized vial with 1ml of Standard Diluent (1X) to generate a **1 ug/ml Standard Solution**. Keep the vial for 15 mins with gentle agitation before making further dilutions. Prepare further **Standards** by diluting the Standard Solution as per the below table. Use the Standard Diluent (1X) as the Zero Standard (Standard No.0).

Standard Concentration	Standard Vial	Dilution Particulars
1 ug/ml	Lyophilized Standard	Original Standard provided in the Kit + 1ml of Standard Diluent (1X)
20 ng/ml	Standard No.6	20 ul Original Standard + 980 ul Standard Diluent (1X)
10 ng/ml	Standard No.5	500 ul Standard No.6 + 500 ul Standard Diluent (1X)
5 ng/ml	Standard No.4	500 ul Standard No.5 + 500 ul Standard Diluent (1X)

2.5 ng/ml	Standard No.3	500 ul Standard No.4 + 500 ul Standard Diluent (1X)
1.25 ng/ml	Standard No.2	500 ul Standard No.3 + 500 ul Standard Diluent (1X)
0.625 ng/ml	Standard No.1	500 ul Standard No.2 + 500 ul Standard Diluent (1X)
0 ng/ml	Standard No.0	Only Standard Diluent (1X)

5. Detection antibody working solution – Reconstitute the lyophilized detection with 150ul of Assay Diluent (1X) to get 3ug/ml. To make 35ng/ml working stock of detection antibody add 58.33ul of detection antibody to 4941.67ul of Assay Diluent (1X) to make final volume to 5ml.
6. Streptavidin HRP working solution – Add 10.87ul of Strep HRP to 4989.13ul of Assay Diluent (1X) to make final volume to 5ml of Streptavidin HRP working solution

Procedural Notes:

1. In order to achieve good assay reproducibility and sensitivity, proper washing of the plates to remove excess un-reacted reagents is essential.
2. High Dose Hook Effect may be observed in samples with very high concentrations of Darbepoetin. High Dose Hook Effect is due to excess of antibody for very high concentrations of Darbepoetin present in the sample. High Dose Hook effect is most likely encountered from samples early in the purification process. If Hook Effect is possible, the samples to be assayed should be diluted with a compatible diluent. Thus if the Darbapoetin concentration of the undiluted sample is less than the diluted sample, this may be indicative of the Hook Effect.
3. Avoid assay of Samples containing sodium azide (NaN_3), as it could destroy the HRP activity resulting in under-estimation of the amount of Darbepoetin.
4. It is recommended that all Standards and Samples be assayed in duplicates.
5. Maintain a repetitive timing sequence from well to well for all the steps to ensure that the incubation timings are same for each well.
6. If the Substrate has a distinct blue color prior to use it may have been contaminated and use of such substrate can lead to compromisation of the sensitivity of the assay.
7. The plates should be read within 30 minutes after adding the Stop Solution.
8. Make a work list in order to identify the location of Standards and Samples.

Assay Procedure:

1. It is strongly recommended that all Standards and Samples be run in duplicates or triplicates. A standard curve is required for each assay. All steps must be performed at 37°C

2. Pipette out **50 ul Assay Diluent** into the respective wells.

3. Pipette **100 ul** of prepared **Standards** or diluted **Samples** into the respective wells.

4. Cover the plate and incubate for 120 minutes at 37°C

5. Aspirate and wash plate 4 times with **Wash Buffer (1X)** and blot residual buffer by firmly tapping plate upside down on absorbent paper. Wipe of any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.

6. Add **100 ul** of **Darbepoetin detection antibody working solution** into each well.

7. Cover the plate and incubate for 60 minutes at 37°C

8. Aspirate and wash plate 4 times with **Wash Buffer (1X)** and blot residual buffer by firmly tapping plate upside down on absorbent paper. Wipe of any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.

9. Add **100 ul** of **Streptavidin HRP working solution** into each well.

10. Cover the plate and incubate for 30 minutes at 37°C.

11. Aspirate and wash plate 4 times with **Wash Buffer (1X)** and blot residual buffer by firmly tapping plate upside down on absorbent paper. Wipe of any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.

12. Add **100 ul** of **TMB Substrate** in each well.

13. Incubate the plate at 37°C for 30 minutes in dark. DO NOT SHAKE or else it may result in higher backgrounds and worse precision. Positive wells should turn bluish in color.

14. Pipette out **100 ul** of **Stop Solution**. Wells should turn from blue to yellow in color.

15. Read the absorbance at 450 nm with a microplate reader.

Calculation of Results:

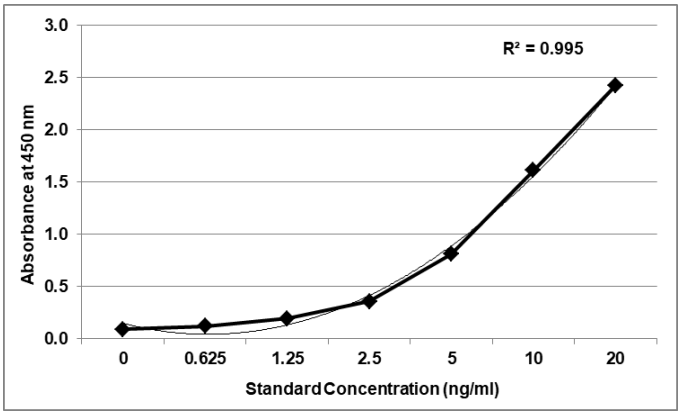
Determine the Mean Absorbance for each set of duplicate or triplicate Standards and Samples. Using Semi-Log graph paper, plot the average value (absorbance 450nm) of each standard on the Y-axis versus the corresponding concentration of the standards on the X-axis. Draw the best fit curve through the standard points. To determine the unknown Darbepoetin concentrations, find the unknown's Mean Absorbance value on the Y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the Darbepoetin Concentration. If samples were diluted, multiply by the appropriate dilution factor. Software which is able to generate a cubic spline curve-fit is best recommended for automated results.

Note:
It is recommended to repeat the assay at a different dilution factor in the following cases:
- If the sample absorbance value is below the first standard.
- If the absorbance value is equivalent or higher than the 20 ng/ml standard.

Typical Data

Standard Concentration (ng/ml)	Abs A	Abs B	Mean Abs	Interpolated Concentration	% Interpolated Concentration against Actual Concentration
0	0.091	0.088	0.090	--	--
0.625	0.113	0.127	0.120	0.6	85.1
1.25	0.208	0.186	0.197	1.4	107.5
2.5	0.358	0.355	0.357	2.5	98.6
5	0.783	0.841	0.812	5.0	100.0
10	1.664	1.563	1.614	10.0	100.1
20	2.404	2.445	2.425	20.0	100.0

Typical Graph



Abs = absorbance @450nm

Quality Control:

It is recommended that for each laboratory assay appropriate quality control samples in each run to be used to ensure that all reagents and procedures are correct.

Performance Characteristics of the Kit:

This kit has been validated as per EMA/FDA guidelines in line with ICH Code for Harmonization of Biological Assays.

Sensitivity:
Limit Of Detection: It is defined as the lowest detectable concentration corresponding to a signal of Mean of '0' standard plus 2* SD.
10 replicates of '0' standards were evaluated and the LOD was found to be less than 0.6 ng/ml.

Precision:

Precision is defined as the percent coefficient of variation (%CV) i.e. standard deviation divided by the mean and multiplied by 100. Assay precision was determined by both intra (n=5 assays) and inter assay (n=5 assays) reproducibility on two pools with low (0.625ng/ml), medium (5ng/ml) and high (20ng/ml) concentrations. While actual precision may vary from laboratory to laboratory and technician to technician, it is recommended that all operators achieve precision below these design goals before reporting results.

Pool	Intra Assay %CV	Inter Assay %CV
Low	<10%	<10%
Medium	<5%	<5%
High	<5%	<5%

Safety Precautions:

- **This kit is For Research Use only**. Follow the working instructions carefully.
- The expiration dates stated on the kit are to be observed. The same relates to the stability stated for reagents
- Do not use or mix reagents from different lots.
- Do not use reagents from other manufacturers.
- Avoid time shift during pipetting of reagents.
- All reagents should be kept in the original shipping container.
- Some of the reagents contain small amount of sodium azide (< 0.1 % w/w) as preservative. They must not be swallowed or allowed to come into contact with skin or mucosa.
- Source materials maybe derived from human body fluids or organs used in the preparation of this kit were tested and found negative for HBsAg and HIV as well as for HCV antibodies. However, no known test guarantees the absence of such viral agents. Therefore, handle all components and all patient samples as if potentially hazardous.
- Since the kit contains potentially hazardous materials, the following precautions should be observed
  - Do not smoke, eat or drink while handling kit material
  - Always use protective gloves
  - Never pipette material by mouth
  - Wipe up spills promptly, washing the affected surface thoroughly with a decontaminant.
- In any case GLP should be applied with all general and individual regulations to the use of this kit.

SCHEMATIC ASSAY PROCEDURE

1. Remove all components, 30 minutes before adding into the assay plate.

Components (2-8ºC) → 30mins RT Before Use → 

→ Thaw at Room Temperature (18-24ºC) → USE NOW

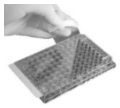
2. Avoid repeated cool-thaw of the components as there will be a loss of activity and this can affect the results.

Kit Components → AVOID → COOL (2-8ºC) → THAW (22±4ºC) → AVOID → NO CHANGE IN RESULTS

THAW (22±4ºC) → COOL (2-8ºC)

3.  Pipette **50 ul Assay diluent** into the respective wells.

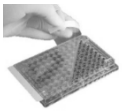
4.  Pipette **100 ul Standards** / **Samples** into the respective wells

5. Cover plate  and incubate for 

at 37°C.

6.  Aspirate and wash wells 4 times with **Wash Buffer (1X)**

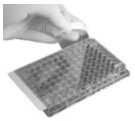
7.  Pipette **100 ul** of **Detection antibody** into the respective wells.

8. Cover plate  and incubate for 

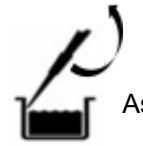
at 37°C.

9.  Aspirate and wash wells 4 times with **Wash Buffer (1X)**

10.  Pipette **100 ul** of **HRP conjugate** into each well.

11. Cover plate  and incubate for 

at 37°C.

12.  Aspirate and wash wells 4 times with **Wash Buffer (1X).**

13.  Pipette **100 ul TMB Substrate** into each well.




14. Cover plate and incubate for at 37°C

15.  Pipette **100 ul Stop Solution** into each well.

16. Read absorbance at 450nm with a  microplate reader within  of stopping reaction.

Typical Example of a Work List

Well #	Contents	Absorbance at 450nm	Mean Absorbance	ng/ml Darbepoetin equivalent
1A 2A	Zero Std Zero Std			
1B 2B	0.625 ng/ml 0.625 ng/ml			
1C 2C	1.25 ng/ml 1.25 ng/ml			
1D 2D	2.5 ng/ml 2.5 ng/ml			
1E 2E	5 ng/ml 5 ng/ml			
1F 2F	10 ng/ml 10 ng/ml			
1G 2G	20 ng/ml 20 ng/ml			
1H 2H	Sample			
3A 4A	Sample			
3B 4B	Sample			

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KRIBIOLISA™ Darbepoetin (ARANESP) ELISA

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KRISHGEN BIOSYSTEMS | OUR REAGENTS | YOUR RESEARCH |

	A MTP epoetin Coated Microtiter Plate (12x8 wells)
	D STD tin Standard
	STRP HRP HRP
	DETN AB detection Antibody
	1X ASY DIL uent
	1X STD DIL Diluent
	1X SAMP DIL luent
	20X WASH BUF er
	SUB TMB ate
	SOLN STOP
	Consult Instructions for Use
	C REF FE umber
	Expiration Date
	Storage Temperature

SYMBOLS KEY