

KRIBIOLISA™ Chrysalin (PT508) ELISA

REF : KBI5005

Ver.1.0

RUO

Enzyme Immunoassay for the Quantitative Determination of
Chrysalin in human serum and plasma

RUO	For Research Use Only	REF	Catalog Number
	Store At	LOT	Batch Code
	Manufactured By		Biological Risk
	Expiry Date		Consult Operating Instructions

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REF KBI5005

 96 tests

KRIBIOLISA™ Chrysalin (PT508) ELISA**Introduction:**

Chrysalin (Rusalatide Acetate), Thrombin Receptor Activating Peptide; TP 508; TRAP-5080 was initially developed to accelerate healing of dermal and musculoskeletal injuries. It proved to be especially effective in promoting healing of diabetic foot ulcers in early human clinical trials and is under development as a treatment for mass casualty injuries and burns. Novel regenerative peptide TP508 also mitigates radiation-induced gastrointestinal damage by activating stem cells and preserving crypt integrity. It is a twenty-three-amino-acid peptide that represents a portion of the receptor-binding domain of the native human thrombin molecule that has been identified as the binding site for a specific class of receptors on fibroblasts and other cells. Novel drugs based on TP508 are under development.

Intended Use:

The KRIBIOLISA™ Chrysalin (PT508) ELISA is used as an analytical tool for quantitative determination of Chrysalin in human serum and plasma.

Principle:

The Chrysalin ELISA is a competitive immunoassay for the determination of Rusalatide Acetate. Plates are coated with Thrombin receptor antibody. A varying concentration of unlabeled standard or sample and constant concentration of Rusalatide Acetate: HRP conjugate will bind in sequence to the antibodies coated on the microplate. Upon washing, unbound Rusalatide Acetate:HRP conjugate will be removed. Bound Rusalatide Acetate:HRP conjugate complex will produce a soluble blue colored product after the addition of TMB Substrate. The enzyme reaction is stopped by dispensing of stop solution into the wells. The optical density (OD) of the solution at 450 nm is inversely proportional to the amount of bound Chrysalin present in the standards or samples.

Materials Provided:

Part	Description	Qty
Thrombin receptor antibody Coated Microtiter Plate	96 well polystyrene microplate (12 strips of 8 wells) coated with Thrombin receptor antibody Coated Microtiter Plate	1 x 96 wells
Rusalatide Acetate standard	Rusalatide Acetate standard (lyophilized, concentration - 1000 ng/ml)	2 vials
Rusalatide Acetate:HRP Conjugate	Rusalatide Acetate:HRP Conjugate prepared in buffer with protein stabilizer and preservatives 0.02% methylisothiazolone and 0.02% bromonitrodioxane.	12 ml
(1X) Sample Diluent	Buffered protein base with preservative 0.02% methylisothiazolone and 0.02% bromonitrodioxane	50 ml
(1X) Standard Diluent	Buffered protein base with protein stabilizer and preservatives 0.02% methylisothiazolone and 0.02% bromonitrodioxane with 1:10 dilution human serum	10 ml
(20X) Wash Buffer	20-fold concentrated solution of buffered surfactant with preservative thiomersal < 0.01%. May turn yellow over time.	25 ml
TMB Substrate	Stabilized Chromogen	12 ml
Stop Solution	0.73M Phosphoric Acid	12 ml
Instruction Manual		1 no

Materials to be provided by the End-User:

1. Microplate Reader able to measure absorbance at 450 nm.
2. Adjustable pipettes to measure volumes ranging from 50 ul to 1000 ul.
3. Deionized (DI) water.
4. Wash bottle or automated microplate washer.
5. Graph paper or software for data analysis.
6. Tubes to prepare standard/sample dilutions.
7. Timer.
8. Absorbent paper.
9. Incubator

KRIBIOLISA™ Chrysalin (PT508) ELISA**Handling/Storage:**

1. All reagents should be stored at 2°C to 8°C for stability.
2. All the reagents and wash solutions should be used within 12 months from manufacturing date.
3. Before using, bring all components to room temperature (18-25°C). Upon assay completion ensure all components of the kit are returned to appropriate storage conditions.
4. The Substrate is light-sensitive and should be protected from direct sunlight or UV sources.

Health Hazard Warnings:

1. Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin.
2. For Research Use Only.

**Sample Preparation and Storage:**

Blood is taken by venipuncture. Serum is separated after clotting by centrifugation. Plasma can be used, too. Lipaemic, hemolytic or contaminated samples should not be run. Repeated freezing and thawing should be avoided. If samples are to be used for several assays, initially aliquot samples and keep at -20°C.

For Cell Culture Supernatant – If necessary, centrifuge to remove debris prior to analysis. Samples can be stored at -20°C or -80°C. Avoid repeated freeze-thaw cycles.

For Serum - Samples have to be diluted 1:10 (v/v), e.g. 1 ul sample + 9 ul sample diluent prior to assay. The samples may be kept at 2 - 8°C for up to three days. Long-term storage requires -20°C.

For Plasma - Samples have to be diluted 1:10 (v/v), e.g. 1 ul sample + 9 ul sample diluent prior to assay. The samples may be kept at 2 - 8°C for up to three days. Long-term storage requires -20°C.

Preparation Before Use:

Allow samples to reach room temperature prior to assay. Take care to agitate patient samples gently in order to ensure homogeneity.

Reagent Preparation (all reagents should be diluted immediately prior to use):

1. Label any aliquots made with the kit Lot No and Expiration date and store it at appropriate conditions mentioned.
2. Bring all reagents to Room Temperature before use.
3. To make **(1X) Wash Buffer**; dilute **25 ml of 20X Wash Buffer in 475 ml of DI water**.
4. **Standards Preparation:** Reconstitute the concentrated Standard lyophilized vial with 1 ml of Standard Diluent to obtain 1000 ng/ml. Keep the vial for 15 mins with gentle agitation and then run the assay procedure. Use the Standard Diluent as the zero standard. Below table shows the calculation for the standard range.

Standard Concentration (ng/ml)	Standard No.	Dilution Particulars
1000 ng/ml	Lyophilized standard	Lyophilized Standard + 1ml of Standard Diluent
31.2 ng/ml	Standard No.5	31.2 ul Reconstituted Standard +968.8 ul Standard Diluent (1X)
15.6 ng/ml	Standard No.4	500ul Standard No.5 + 500ul Standard Diluent (1X)
7.8 ng/ml	Standard No.3	500ul Standard No.4 + 500ul Standard Diluent (1X)
3.9 ng/ml	Standard No.2	500ul Standard No.3 + 500ul Standard Diluent (1X)
1.95 ng/ml	Standard No.1	500ul Standard No.2 + 500ul Standard Diluent (1X)
0	Standard No.0	Only Standard Diluent (1X)

Mix each tube thoroughly before the next transfer. Use the standards for experiment within one hour of preparation of standard. Discard standard after use.

KRIBIOLISA™ Chrysalin (PT508) ELISA**Procedural Notes:**

1. In order to achieve good assay reproducibility and sensitivity, proper washing of the plates to remove excess un-reacted reagents is essential.
2. Avoid assay of Samples containing Sodium Azide (NaN_3), as it could destroy the HRP activity of the conjugate resulting in under-estimation of the antibodies.
3. It is recommended that all Standards and Samples be assayed in duplicates or triplicates.
4. Maintain a repetitive timing sequence from well to well for all the steps to ensure that the incubation timings are same for each well.
5. If the Substrate has a distinct blue color prior to use it may have been contaminated and use of such substrate can lead to compromise of the sensitivity of the assay.
6. The plates should be read within 30 minutes after adding the Stop Solution.
7. Make a work list in order to identify the location of Standards and Samples.
8. Making serial dilution in the wells directly is not permitted.
9. Prepare the Standard within 15 minutes prior to running the assay.
10. Please carefully dilute Standards according to the instruction, and avoid foaming. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettes are calibrated.
11. If crystals have formed in the Wash Solution (20X) concentrate, warm to room temperature and mix gently until the crystals are completely dissolved.
12. Contaminated water or container for reagent preparation will influence the detection results.

Assay Procedure:

1. It is strongly recommended that all Standards and Samples be run in duplicates or triplicates. A standard curve is required for each assay. All steps must be performed at RT.
2. Pipette out **100 ul of Standards and samples** to the respective wells.
3. Add **100 ul Rusalatide Acetate:HRP Conjugate** to each well.
4. Cover the plate and incubate for 210 mins at 37°C.
5. Aspirate and wash plate 4 times with **Wash Buffer (1X)** and blot residual buffer by firmly tapping plate upside down on absorbent paper. Wipe of any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.
6. Add **100 ul of TMB Substrate** in each well.
7. Incubate the plate at RT for 30 minutes in dark. DO NOT SHAKE or else it may result in higher backgrounds and worse precision. Positive wells should turn bluish in color.
8. Pipette out **100 ul of Stop Solution**. Wells should turn from blue to yellow in color.
9. Read the absorbance at 450 nm with a microplate reader.

Calculation of Results:

Determine the Mean Absorbance for each set of duplicate or triplicate Standards and Samples. Using Graph paper, plot the average value (absorbance 450nm) of each standard on the Y-axis versus the corresponding concentration of the standards on the X-axis. Draw the best fit curve through the standard points. To determine the unknown Chrysalin concentrations, find the unknown's Mean Absorbance value on the Y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the Chrysalin Concentration. If samples were diluted, multiply by the appropriate dilution factor. Software which is able to generate a cubic spline curve-fit is best recommended for automated results.

Note:

It is recommended to repeat the assay at a different dilution factor in the following cases:

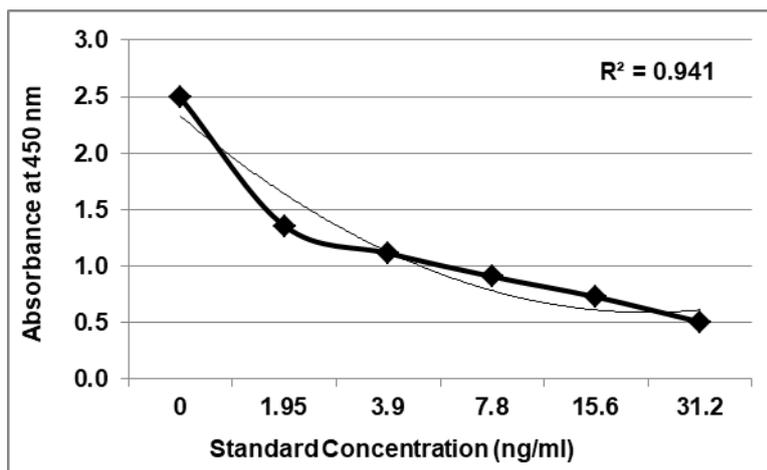
- If the sample absorbance value is below the first standard.
- If the absorbance value is equivalent or higher than the 31.2 ng/ml standard.

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Typical Data

Standard Concentration (ng/ml)	Mean Absorbance	Interpolated Concentration	% Interpolated Concentration against Actual Concentration
0	2.490	0.000	--
1.95	1.356	1.9	96.7
3.9	1.113	4.1	105.4
7.8	0.906	7.9	101.8
15.6	0.726	14.5	92.8
31.2	0.505	32.4	104.0

Typical Graph



abs = absorbance at 450nm

Quality Control:

It is recommended that for each laboratory assay appropriate quality control samples in each run to be used to ensure that all reagents and procedures are correct.

Performance Characteristics:

Please note that this validation is performed in our laboratory and will not necessarily be duplicated in your laboratory. This data has been generated to enable the user to get a preview of the assay and the characteristics of the kit and is generic in nature. We recommend that the user performs at the minimum; the spike and recovery assay and the dilutional linearity assay to assure quality results. For a more comprehensive validation, the user may run the protocols as suggested by us herein below to develop the parameters for quality control to be used with the kit.

Detection Range:

1.95ng/ml – 31.2ng/ml.

Precision:

Intra-Assay Precision: 3 samples with low, middle and high level Chrysalin were tested 20 times on one plate, respectively.

Inter-Assay Precision: 3 samples with low, middle and high level Chrysalin were tested on 3 different plates, 8 replicates in each plate.

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CV (%) = SD/mean X 100

Intra-Assay: CV<15%

Inter-Assay: CV<18%

Safety Precautions:

- **This kit is For Research Use only.** Follow the working instructions carefully.
- The expiration dates stated on the kit are to be observed. The same relates to the stability stated for reagents
- Do not use or mix reagents from different lots.
- Do not use reagents from other manufacturers.
- Avoid time shift during pipetting of reagents.
- All reagents should be kept in the original shipping container.
- Some of the reagents contain small amount of sodium azide (< 0.1 % w/w) as preservative. They must not be swallowed or allowed to come into contact with skin or mucosa.
- Source materials maybe derived from human body fluids or organs used in the preparation of this kit were tested and found negative for HBsAg and HIV as well as for HCV antibodies. However, no known test guarantees the absence of such viral agents. Therefore, handle all components and all patient samples as if potentially hazardous.
- Since the kit contains potentially hazardous materials, the following precautions should be observed
 - Do not smoke, eat or drink while handling kit material
 - Always use protective gloves
 - Never pipette material by mouth
 - Wipe up spills promptly, washing the affected surface thoroughly with a decontaminant.
- In any case GLP should be applied with all general and individual regulations to the use of this kit.



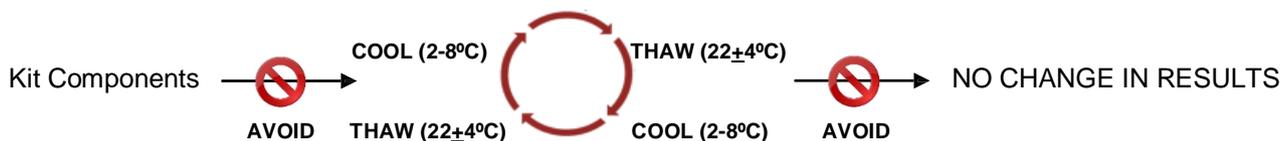
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SCHEMATIC ASSAY PROCEDURE

1. Remove all components, 30 minutes before adding into the assay plate.



2. Avoid repeated cool-thaw of the components as there will be a loss of activity and this can affect the results.



3.  Pipette **100 ul Standard and Sample** into respective well.

4.  Pipette **100 ul Rusalatide Acetate:HRP Conjugate** into the respective wells.

5. Cover plate  and incubate for  at 37°C.

6.  Aspirate and wash wells **4 times** with **Wash Buffer (1X)**.

7.  Pipette **100 ul TMB Substrate** into each well.

8. Cover plate  and incubate for  at RT.

9. Read absorbance at 450nm with a  microplate reader within  of stopping reaction.

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Typical Example of a Work List

Well #	Contents	Absorbance at 450nm	Mean Absorbance	ng/ml Rusalatide
1A	0 ng/ml			
2A	0 ng/ml			
1B	1.95 ng/ml			
2B	1.95 ng/ml			
1C	3.9 ng/ml			
2C	3.9 ng/ml			
1D	7.8 ng/ml			
2D	7.8 ng/ml			
1E	15.6 ng/ml			
2E	15.6 ng/ml			
1F	31.2 ng/ml			
2F	31.2 ng/ml			
1G	Sample			
2G				
1H	Sample			
2H				
3A	Sample			
4A				
3B	Sample			
4B				

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SYMBOLS KEY

	Thrombin receptor antibody Coated Microtiter Plate (12x8 wells)
	Standard, lyophilized
	Conjugate Horseradish Peroxidase
	(1X) Sample Diluent
	(1X) Standard Diluent
	(20X) Wash Buffer
	TMB Substrate
	Stop Solution
	Consult Instructions for Use
	Catalog Number
	Expiration Date
	Storage Temperature