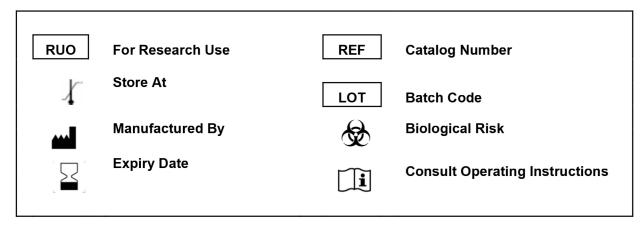
KRIBIOLISA™ Eculizumab (Soliris™) ELISA

: KBI1024 Ver 4.0

RUO

Enzyme Immunoassay for the quantitative determination of Eculizumab in serum, plasma and cell culture supernatant



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Introduction:

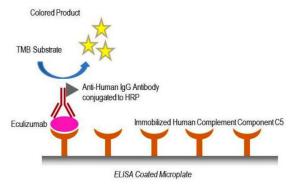
Eculizumab, sold under the trade name Soliris, is a medication used to treat paroxysmal nocturnal hemoglobinuria (PNH) and atypical hemolytic uremic syndrome (aHUS). It is a humanized monoclonal antibody functioning as a terminal complement inhibitor. In people with PNH, it reduces both the destruction of red blood cells and need for blood transfusion, but does not appear to affect the risk of death. Eculizumab was the first drug approved for each of its uses, and its approval was granted on the basis of small clinical trials.

Intended Use:

The KRIBIOLISA™ Eculizumab (Soliris™) ELISA is used as an analytical tool for quantitative determination of Eculizumab in human serum and plasma

Principle:

The method employs the quantitative sandwich enzyme immunoassay technique. Human complement component C5 protein is pre-coated onto microwells. Samples and standards are pipetted into microwells and human Eculizumab present in the sample are bound by the capture protein. Then, a HRP (horseradish peroxidase) conjugated anti-Human IgG antibody is pipetted and incubated. After washing microwells in order to remove any non-specific binding, the ready to use substrate solution (TMB) is added to microwells and color develops proportionally to the amount of Eculizumab in the sample. Color development is then stopped by addition of stop solution. Absorbance is measured at 450 nm.



Materials Provided:

| Part | Description | Qty |
|---|---|--------------|
| Human complement component C5 Coated Microtiter Plate | 96 well polystyrene microplate (12 strips of 8 wells) coated with purified human complement component C5 | 1 x 96 wells |
| Eculizumab Standard | Recombinant Anti-C5 Antibody (Eculizumab) IgG2/4 kappa antibody in a buffered protein base and preservative sodium azide < 0.01% – lyophilized (1 ug/ml) | 2 vials |
| Anti-Human IgG:HRP Conjugate | Goat Anti-Human IgG conjugated to Horseradish Peroxidase with protein stabilizer and preservatives 0.02% methylisothiazolone and 0.02% bromonitrodioxane. | 12 ml |
| Standard Diluent | Buffered protein base with 1:1000 dilution human serum and preservative sodium azide < 0.01% | 10 ml |
| (1X) Sample Diluent | Buffered protein base with preservative thiomersol < 0.01% | 2 x 50 ml |
| (20X) Wash Buffer | 20-fold concentrated solution of buffered surfactant with preservative thiomersol < 0.01%. May turn yellow over time. | 25 ml |
| TMB Substrate | Stabilized Chromogen | 12 ml |
| Stop Solution | 2N Sulfuric Acid | 12 ml |
| Instruction Manual | | 1 no |

Materials to be provided by the End-User:

1. Microtiter Plate Reader able to measure absorbance at 450 nm.

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- 2. Adjustable pipettes and multichannel pipettor to measure volumes ranging from 25 ul to 1000 ul
- 3. Deionized (DI) water
- 4. Wash bottle or automated microplate washer
- 5. Graph paper or software for data analysis
- 6. Timer
- 7. Absorbent Paper

Handling/Storage:

- 1. All reagents should be stored at 2°C to 8°C for stability.
- 2. All the reagents and wash solutions should be used within 12 months from manufacturing date.
- 3. Before using, bring all components to room temperature (18-25°C). Upon assay completion ensure all components of the kit are returned to appropriate storage conditions.
- 4. The Substrate is light-sensitive and should be protected from direct sunlight or UV sources.

Health Hazard Warnings:

- 1. Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin.
- 2. For Research Use Only.



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Sample Preparation and Storage:

Blood is taken by venipuncture. Serum is separated after clotting by centrifugation. Plasma can be used, too. Lipaemic, hemolytic or contaminated samples should not be run. Repeated freezing and thawing should be avoided. If samples are to be used for several assays, initially aliquot samples and keep at -20°C.

For Serum / Plasma - Samples have to be diluted 1:1000 (v/v), e.g. 1 ul sample + 999 ul (1X) sample diluent prior to assay. The samples may be kept at $2 - 8^{\circ}$ C for up to three days. Long-term storage requires -20°C.

Reagent Preparation (all reagents should be diluted immediately prior to use):

- Label any aliquots made with the kit Lot No and Expiration date and store it at appropriate conditions mentioned.
- 2. Bring all reagents to Room temperature before use.
- 3. To make Wash Buffer (1X); dilute 25 ml of 20X Wash Buffer in 475 ml of DI water.
- 4. Standards Preparation: Reconstitute the concentrated Standard lyophilized vial with 1 ml of Standard Diluent to obtain a concentration of 1 ug/ml. Keep the vial for 15 mins with gentle agitation before making further dilutions. Dilute 320 ul of original Standard (1 ug/ml) with 180 ul of Standard Diluent to generate a 640 ng/ml Standard Solution. Prepare further Standards by serially diluting the Standard Solution as per the below table. Use the Standard Diluent as the Zero Standard (Standard No.0).

| Standard Concentration | Standard Vial | Dilution Particulars |
|------------------------|-------------------------|---|
| 1 ug/ml | Lyophilized Standard | Lyophilized Standard provided in the Kit + 1ml of Standard Diluent |
| 640 ng/ml | Standard No.7 | 320 ul Reconstituted Standard (1 ug/ml) + 180 ul Standard Diluent |
| 320 ng/ml | Standard No.6 | 250 ul Standard No.7 + 250 ul Standard Diluent |
| 160 ng/ml | Standard No.5 | 250 ul Standard No.6 + 250 ul Standard Diluent |
| 80 ng/ml | Standard No.4 | 250 ul Standard No.5 + 250 ul Standard Diluent |
| 40 ng/ml | Standard No.3 | 250 ul Standard No.4 + 250 ul Standard Diluent |
| 20 ng/ml | Standard No.2 | 250 ul Standard No.3 + 250 ul Standard Diluent |
| 10 ng/ml | Standard No.1 | 250 ul Standard No.2 + 250 ul Standard Diluent |
| 0 ng/ml | Standard No.0 | Only Standard Diluent |



Use the Standards immediately upon reconstitution. Discard balance standard after use. Do not store them for further experiments.

Procedural Notes:

- 1. In order to achieve good assay reproducibility and sensitivity, proper washing of the plates to remove excess un-reacted reagents is essential.
- 2. High Dose Hook Effect may be observed in samples with very high concentrations of Eculizumab. High Dose Hook Effect is due to excess of antibody for very high concentrations of Eculizumab present in the sample. High Dose Hook effect is most likely encountered from samples early in the purification process. If Hook Effect is possible, the samples to be assayed should be diluted with a compatible diluent. Thus if the Eculizumab concentration of the undiluted sample is less than the diluted sample, this may be indicative of the Hook Effect.
- 3. Avoid assay of Samples containing sodium azide (NaN₃), as it could destroy the HRP activity resulting in under-estimation of the amount of Eculizumab.
- 4. It is recommended that all Standards and Samples be assayed in duplicates.
- 5. Maintain a repetitive timing sequence from well to well for all the steps to ensure that the incubation timings are same for each well.
- 6. If the Substrate has a distinct blue color prior to use it may have been contaminated and use of such substrate can lead to compromisation of the sensitivity of the assay.
- 7. The plates should be read within 30 minutes after adding the Stop Solution.
- 8. Make a work list in order to identify the location of Standards and Samples.

Assay Procedure:

- 1. It is strongly recommended that all Controls and Samples be run in duplicates. A standard curve is required for each assay. All steps must be performed at 37°C
- 2. Add 100 ul of Standards or Samples into the respective wells.
- 3. Cover the plate and incubate for 60 minutes at 37°C
- 4. Aspirate and wash plate 4 times with Wash Buffer (1X) and blot residual buffer by firmly tapping plate upside down on absorbent paper. Wipe of any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.
- 5. Pipette without delay in the same order 100 ul of Anti-Human IgG:HRP Conjugate into each well.
- 6. Cover the plate and incubate for 60 minutes at 37°C
- 7. Aspirate and wash plate 4 times with Wash Buffer (1X) and blot residual buffer by firmly tapping plate upside down on absorbent paper. Wipe of any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.
- 8. Add 100 ul of TMB Substrate in each well.
- 9. Incubate the plate at 37°C for 30 minutes in dark. DO NOT SHAKE or else it may result in higher backgrounds and worse precision. Positive wells should turn bluish in color.
- 10. Pipette out 100 ul of Stop Solution. Wells should turn from blue to yellow in color.
- 11. Read the absorbance at 450 nm with a microplate reader.

Calculation of Results:

Determine the Mean Absorbance for each set of duplicate Standards and Samples. Using Graph paper, plot the average value (absorbance 450nm) of each standard on the Y-axis versus the corresponding concentration of the standards on the X-axis. Draw the best fit curve through the standard points.

To determine the unknown Eculizumab concentrations, find the unknown's Mean Absorbance value on the Y-4



axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the Eculizumab Concentration.

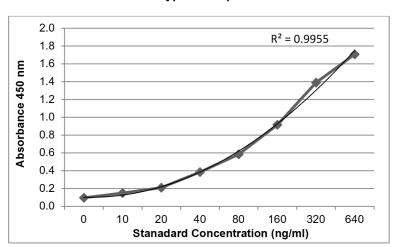
If samples were diluted, multiply by the appropriate dilution factor.

Software which is able to generate a cubic spline curve-fit or a polynomial curve (2nd order) is best recommended for automated results.

Note:

It is recommended to repeat the assay at a different dilution factor in the following cases:

- If the sample absorbance value is below the first standard.
- If the absorbance value is equivalent or higher than the 640 ng/ml standard.



Typical Graph

Quality Control:

It is recommended that for each laboratory assay appropriate quality control samples in each run to be used to ensure that all reagents and procedures are correct.

Performance Characteristics of the Kit:

This kit has been validated as per EMA/FDA guidelines in line with ICH Code for Harmonization of Biological Assays.

Sensitivity:

Limit Of Detection: It is defined as the lowest detectable concentration corresponding to a signal of Mean of '0' standard plus 2* SD.

10 replicates of '0' standards were evaluated and the LOD was found to be less than 8 ng/ml

Specificity:

The protein used in the kit is purified C5 and binds to Eculizumab. The calibrators / standards used are calibrated against commercially sourced Soliris $^{\text{TM}}$.

Precision:

Precision is defined as the percent coefficient of variation (%CV) i.e. standard deviation divided by the mean and multiplied by 100. Assay precision was determined by both intra (n=5 assays) and inter assay (n=5 assays) reproducibility on two pools with low (10 ng/ml), medium (80 ng/ml) and high (640 ng/ml) concentrations.

While actual precision may vary from laboratory to laboratory and technician to technician, it is recommended that all operators achieve precision below these design goals before reporting results.

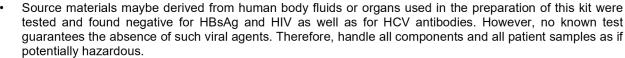
| Pool | Intra Assay %CV | Inter Assay %CV |
|------|-----------------|-----------------|
| Low | <10% | <10% |



| Medium | <5% | <5% |
|--------|-----|-----|
| High | <5% | <5% |

Safety Precautions:

- This kit is for Research use only. Follow the working instructions carefully.
- The expiration dates stated on the kit are to be observed. The same relates to the stability stated for reagents
- Do not use or mix reagents from different lots.
- Do not use reagents from other manufacturers.
- Avoid time shift during pipetting of reagents.
- All reagents should be kept in the original shipping container.
- Some of the reagents contain small amount of sodium azide (< 0.1 % w/w) as preservative. They must not be swallowed or allowed to come into contact with skin or mucosa.





- Do not smoke, eat or drink while handling kit material
- Always use protective gloves
- Never pipette material by mouth
- Wipe up spills promptly, washing the affected surface thoroughly with a decontaminant.
- In any case GLP should be applied with all general and individual regulations to the use of this kit.

References:

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Variable Eculizumab Clearance Requires Pharmacodynamic Monitoring to Optimize Therapy for Thrombotic Microangiopathy after Hematopoietic Stem Cell Transplantation...Jodele S, Fukuda T, Mizuno K, Vinks AA, Laskin BL, Goebel J, Dixon BP, Chima RS, Hirsch R, Teusink A, Lazear D, Lane A, Myers KC, Dandoy CE, Davies SM... Biol Blood Marrow Transplant....2016....Elsevier





USE NOW



SCHEMATIC ASSAY PROCEDURE

1. Remove all components, 30 minutes before adding into the assay plate.

Components (2-8°C) 30 mins
RT Before Use



Thaw at Room
Temperature (18-24°C)

2. Avoid repeated cool-thaw of the components as there will be a loss of activity and this can affect the results.



Pipette 100 ul Standards / Samples into the respective wells.

- 4. Cover plate and incubate for **60**min at 37°C
- 5. Aspirate and wash wells 4 times with **Wash Buffer (1X).**
- 6. Pipette 100 ul Anti Human IgG:HRP into each well.
- 7. Cover plate and incubate for 60min at 37°C
- 8. Aspirate and wash wells 4 times with **Wash Buffer (1X).**
- 9. Pipette 100 ul TMB Substrate into each well.
- 10. Cover plate and incubate for **30**min at 37°C
- 11. Pipette 100 ul Stop Solution into each well.
- 12. Read absorbance at 450nm with a microplate reader within **30**min





Typical Example of a Work List

| Well# | Contents | Absorbance at 450nm | Mean Absorbance | ng/ml Eculizumab equivalent |
|----------|------------------------|------------------------|--------------------|-----------------------------------|
| 1A 2A | zero std zero std | | | |
| 1B | 10 ng/ml | | | |
| 2B | 10 ng/ml | | | |
| 1C 2C | 20 ng/ml 20 ng/ml | | | |
| 1D 2D | 40 ng/ml 40 ng/ml | | | |
| 1E 2E | 80 ng/ml 80 ng/ml | | | |
| 1F 2F | 160 ng/ml 160 ng/ml | | | |
| 1G 2G | 320 ng/ml 320 ng/ml | | | |
| 1H 2H | 640 ng/ml 640 ng/ml | | | |
| 3A 4A | Sample | | | |
| 3B 4B | Sample | | | |

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SYMBOLS KEY

| МТР | Microtiter Plate (12X8 wells) |
|--------------|----------------------------------|
| STD | Eculizumab Standard, lyophilized |
| HRP CONJ | Conjugate Horseradish Peroxidase |
| STD DIL | Standard Diluent |
| ASY DIL | Assay Diluent |
| SAMP DIL | Sample Diluent |
| 20X WASH BUF | (20X) Wash Buffer |
| SUB TMB | TMB Substrate |
| SOLN STOP | Stop Solution |
| []i | Consult Instructions for Use |
| REF | Catalog Number |
| \square | Expiration Date |
| * | Storage Temperature |