






Peptidoglycan (PTG) GENLISA™ ELISA

REF : KLU0202

Ver 1.0


RUO

Enzyme Immunoassay for the Quantitative Determination of Peptidoglycan (PTG) in serum, plasma and other biological samples.

RUO	For Research Use Only	REF	Catalog Number
	Store At	LOT	Batch Code
	Manufactured By		Biological Risk
	Expiry Date		Consult Operating Instructions

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 96 tests

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Introduction:

The GENLISA™ ELISA kits are used for assessing the specific biomarker in samples analytes which may be serum, plasma and cell culture supernatant as validated with the kit. The kit employs a competitive ELISA technique.

Intended Use:

The Peptidoglycan (PTG) GENLISA™ ELISA kit is used as an analytical tool for quantitative determination of Peptidoglycan (PTG) in serum, plasma and other biological samples.

Principle:

The method employs competitive enzyme-linked immunosorbent assay (ELISA) technique to assay the level of PTG in samples. Standards or Samples competes with the Biotin labeled antigen to form a complex with the PTG antibody coated microtiter well. After washing Streptavidin:HRP is pipetted and incubated After incubation and a washing step TMB Substrate, are added. Blue color develops on incubation and the reaction is stopped with a Stop Solution to form a yellow color. The concentration of the PTG in the samples is inversely proportional to the yellow color developed (absorbance) in the wells.

Materials Provided:

1. Peptidoglycan (PTG) Antibody Coated Microtiter Plate (12 x 8 wells) – 1 no
2. Standard, Peptidoglycan (PTG) (Lyophilized, 100 ng/ml) – 2 vials
3. Biotinylated Peptidoglycan (PTG) Antigen (concentrated) - 60 ul
4. Streptavidin:HRP Conjugate (concentrated) - 120 ul
5. Sample Diluent - 20 ml
6. Biotin Antibody Dilution Buffer - 13 ml
7. HRP Conjugate Dilution Buffer - 13 ml
8. (20X) Wash Buffer – 25 ml
9. TMB Substrate – 12 ml
10. Stop Solution – 12 ml
11. Instruction Manual

Materials to be provided by the End-User:

1. Microtiter Plate Reader able to measure absorbance at 450 nm.
2. Adjustable pipettes and multichannel pipettor to measure volumes ranging from 25 ul to 1000 ul
3. Deionized (DI) water
4. Wash bottle or automated microplate washer
5. Graph paper or software for data analysis
6. Timer
7. Absorbent Paper

Handling/Storage:

1. All reagents should be stored as indicated on the component label.
2. All the reagents and wash solutions should be used within 12 months from manufacturing date.
3. Before using, bring all components to room temperature (18-25°C). Upon assay completion ensure all components of the kit are returned to appropriate storage conditions.
4. The Substrate is light-sensitive and should be protected from direct sunlight or UV sources.

Health Hazard Warnings:

1. Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin.
2. For Research Use Only.

**Sample Preparation and Storage:**

Specimens should be clear and non-hemolyzed. Samples should be run at a number of dilutions to ensure accurate quantitation.

1. Extract as soon as possible after specimen collection as per relevant procedure. The samples should be tested as soon as possible after the extraction. Alternately the extracted samples can be kept in -20°C. Avoid repeated freeze-thaw cycles.
2. **Serum-** Coagulate at room temperature for 10-20 minutes; centrifuge for 20-min at 2000-3000 rpm. Remove the supernatant. If precipitation appears, recentrifuge.
3. **Plasma-** Use EDTA or citrate plasma as an anticoagulant, mix for 10-20 minutes; centrifuge for 15-min at 2000-3000 rpm. Remove the supernatant carefully. If precipitation appears, recentrifuge.
4. **Urine-** Collect urine in a sterile container, centrifuge for 20-min at 2000-3000 rpm. Remove the supernatant. If precipitation appears, recentrifuge.
5. **Cell Culture Supernatant-** Collect sample in a sterile container. Centrifuge for 20-mins at 2000-3000 rpm. Remove the supernatant carefully. When examining the components within the cell, dilute cell suspension with PBS (pH 7.2-7.4), if cell concentration is greater than 1 million/ml. Damage the cells by repeated freeze-thaw cycles to release intracellular components. Centrifuge for 20-min at 2000-3000 rpm. If precipitation appears, centrifuge again.
6. **Tissue Samples-** Rinse tissues in PBS (pH 7.4) to remove excess blood thoroughly and weigh before homogenization. Mince tissues and homogenize them in PBS (pH7.4) with a glass homogenizer on ice. Thaw at 2-8°C or freeze at -20°C. Centrifuge at 2000-3000 RPM for approximately 20 minutes and collect the supernatant carefully.

Note: Grossly hemolyzed samples are not suitable for use in this assay.

Sample Dilution

Please note the kit is validated for use with neat samples.

Incase the user wishes to estimate the concentration of the target protein in the test sample, we advise to select a proper dilution factor to make the diluted target protein concentration fall in the optimal detection range of the kit and several trials may be necessary. If samples contain very high concentrations of the analyte, dilute the samples with the Sample Diluent provided in the kit.

Reagent Preparation (all reagents should be diluted immediately prior to use):

1. Label any aliquots made with the kit Lot No and Expiration date and store it at appropriate conditions mentioned.
2. Bring all reagents to Room temperature before use.
3. To make **Wash Buffer (1X)**; dilute **25 ml of 20X Wash Buffer in 475 ml of DI water**.
4. **Biotinylated Peptidoglycan (PTG) Antigen Working Solution:** Calculate required total volume of the working solution: 0.05ml / well x quantity of wells. In preparation, slightly more than calculated should be prepared. Centrifuge the stock tube before use, dilute the Biotinylated Peptidoglycan (PTG) Antigen (concentrated) with Biotin Antigen Dilution Buffer at 1:100 and mix them thoroughly (i.e. Add 1 ul Biotinylated Peptidoglycan (PTG) Antigen into 99 ul Biotin Antigen Dilution Buffer.)
5. **Streptavidin:HRP Conjugate Working Solution:** Calculate required total volume of the working solution: 0.1ml / well x quantity of wells. In preparation, slightly more than calculated should be prepared. Centrifuge the stock tube before use, dilute the Streptavidin:HRP Conjugate with Streptavidin:HRP Conjugate Dilution Buffer at 1:100 and mix them thoroughly (i.e. Add 1 ul of Streptavidin:HRP Conjugate into 99 ul of Streptavidin:HRP Conjugate Dilution Buffer).
6. **Standards Preparation:** Reconstitute original Peptidoglycan (PTG) with 1.0 ml of Sample Diluent. Keep the standard for 10 mins with gentle agitation before making further dilutions. Prepare the additional Standards by serially diluting the standard stock solution as per the below table.

Standard Concentration	Standard Vial	Dilution Particulars
100 ng/ml	Standard No.7	Reconstitute with 1.0 ml Sample Diluent
50 ng/ml	Standard No.6	500 ul Standard No.7 + 500 ul Sample Diluent
25 ng/ml	Standard No.5	500 ul Standard No.6 + 500 ul Sample Diluent
12.5 ng/ml	Standard No.4	500 ul Standard No.5 + 500 ul Sample Diluent
6.25 ng/ml	Standard No.3	500 ul Standard No.4 + 500 ul Sample Diluent
3.13 ng/ml	Standard No.2	500 ul Standard No.3 + 500 ul Sample Diluent
1.57 ng/ml	Standard No.1	500 ul Standard No.2 + 500 ul Sample Diluent
0 ng/ml	Standard No.0	50 ul Sample Diluent

Procedural Notes:

1. In order to achieve good assay reproducibility and sensitivity, proper washing of the plates to remove excess un-reacted reagents is essential.
2. High Dose Hook Effect may be observed in samples with very high concentrations of Peptidoglycan (PTG). High Dose Hook Effect is due to excess of antibody for very high concentrations of Peptidoglycan (PTG) present in the sample.
3. Peptidoglycan (PTG) concentration of the undiluted sample is less than the diluted sample, this may be indicative of the Hook Effect.
4. Avoid assay of Samples containing sodium azide (NaN₃), as it could destroy the HRP activity resulting in under-estimation of the amount of Peptidoglycan (PTG).
5. It is recommended that all Standards and Samples be assayed in duplicates or triplicates.
6. Maintain a repetitive timing sequence from well to well for all the steps to ensure that the incubation timings are same for each well.
7. If the Substrate has a distinct blue color prior to use it may have been contaminated and use of such substrate can lead to compromisation of the sensitivity of the assay.
8. The plates should be read within 30 minutes after adding the Stop Solution.
9. Make a work list in order to identify the location of Standards and Samples.

Assay Procedure:

1. It is strongly recommended that all Standards and Samples be run in duplicates or triplicates. A standard curve is required for each assay.
2. Set standard, samples, control (blank) wells on the pre-coated plate respectively, and then, record the positions.
3. Add **50 ul Sample diluent** to the blank wells.
4. Add **50 ul prepared standards** to respective standard wells.
5. Add **50 ul Samples** to the sample wells.
6. Pipette **50 ul Biotinylated Peptidoglycan (PTG) Antigen Working Solution** to all wells.
7. Cover the plate with a sealer and incubate for 60 minutes at 37°C.
8. Aspirate and wash plate 4 times with diluted Wash Buffer (1X) and blot residual buffer by firmly tapping plate upside down on absorbent paper. Wipe of any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.
9. Pipette **100 ul Streptavidin:HRP Conjugate Working Solution** to all wells except the blank wells.
10. Cover the plate with a sealer and incubate for **30 minutes** at **37°C**.
11. Aspirate and wash plate 4 times with diluted **Wash Buffer (1X)** and blot residual buffer by firmly tapping plate upside down on absorbent paper. Wipe of any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.

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12. Pipette **100 ul TMB Substrate** to all wells.
13. Incubate the plate at **37°C** for **15 minutes**. Protect the plate from light.
14. Pipette **100 ul of Stop Solution** to all wells. The wells should turn from blue to yellow in color.
15. Read the absorbance at 450 nm with a microplate within 10-15 minutes after addition of Stop solution.

Calculation of Results:

Determine the Mean Absorbance for each set of duplicate or triplicate Standards and Samples. Using Graph Paper, plot the average value (absorbance 450nm) of each standard on the Y-axis versus the corresponding concentration of the standards on the X-axis. Draw the best fit curve through the standard points. To determine the unknown Peptidoglycan (PTG) concentrations, find the unknown's Mean Absorbance value on the Y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the Peptidoglycan (PTG) Concentration.

If samples were diluted, multiply by the appropriate dilution factor. Software which is able to generate a cubic spline curve-fit or 4-PL is best recommended for automated results.

Note:

It is recommended to repeat the assay at a different dilution factor in the following cases:
- If the sample absorbance value is below the first standard.

Quality Control:

It is recommended that for each laboratory assay appropriate quality control samples in each run to be used to ensure that all reagents and procedures are correct.

Performance Characteristics of the Kit:

This kit has been validated. Please view the details herein below.

Standard Calibration Range:

1.57 ng/ml – 100 ng/ml

Sensitivity:

Limit Of Quantification:

It is defined as the lowest detectable concentration that can be determined with an acceptable repeatability and the LOQ was found to be 0.78 ng/ml.

Specificity:

This assay has high sensitivity and excellent specificity for detection of Peptidoglycan (PTG). No significant cross-reactivity or interference between Peptidoglycan (PTG) and analogues was observed.

Recovery

Matrices listed below were spiked with certain level of Peptidoglycan (PTG) and the recovery rates were calculated by comparing the measured value to the expected amount of Peptidoglycan (PTG) in samples.

Matrix	Recovery Range (%)	Average (%)
serum(n=5)	87-99	93
EDTA plasma(n=5)	83-99	95
cell culture media(n=5)	80-96	88

Precision:

Intra-Assay: CV<10%

Inter-Assay: CV<12%

Linearity

The linearity of the kit was assayed by testing samples spiked with appropriate concentration of Peptidoglycan (PTG) and their serial dilutions. The results were demonstrated by percentage of calculated concentration to the expectation.

Sample	1:2	1:4	1:8	1:16
serum(n=5)	86-96%	93-104%	89-97%	85-94%
EDTA plasma(n=5)	85-92%	85-92%	97-103%	87-98%
cell culture media(n=5)	95-102%	98-103%	86-92%	87-93%

Safety Precautions:

- **This kit is For Research Use only.** Follow the working instructions carefully.
- The expiration dates stated on the kit are to be observed. The same relates to the stability stated for reagents
- Do not use or mix reagents from different lots.
- Do not use reagents from other manufacturers.
- Avoid time shift during pipetting of reagents.
- All reagents should be kept in the original shipping container.
- Some of the reagents contain small amount of sodium azide (< 0.1 % w/w) as preservative. They must not be swallowed or allowed to come into contact with skin or mucosa.
- Source materials maybe derived from body fluids or organs used in the preparation of this kit were tested and found negative for HBsAg and HIV as well as for HCV antibodies. However, no known test guarantees the absence of such viral agents. Therefore, handle all components and all patient samples as if potentially hazardous.
- Since the kit contains potentially hazardous materials, the following precautions should be observed
 - Do not smoke, eat or drink while handling kit material
 - Always use protective gloves
 - Never pipette material by mouth
 - Wipe up spills promptly, washing the affected surface thoroughly with a decontaminant.
- In any case GLP should be applied with all General and individual regulations to the use of this kit.



Typical Example of a Work List

Well #	Contents	Absorbance at 450nm	Mean Absorbance	Interpolated Concentration
1A 2A	Blank Blank			
1B 2B	Standard No.1 Standard No.1			
1C 2C	Standard No.2 Standard No.2			
1D 2D	Standard No.3 Standard No.3			
1E 2E	Standard No.4 Standard No.4			
1F 2F	Standard No.5 Standard No.5			
1G 2G	Standard No.6 Standard No.6			
1H 2H	Standard No.7 Standard No.7			
3A 4A	Sample			
3B 4B	Sample			

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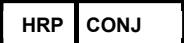

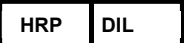








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SYMBOLS KEY

	Coated Microtiter Plate (8x12 wells)
	Standard
	Biotinylated Antigen
	Conjugate Horseradish Peroxidase
	Biotin Antigen Dilution Buffer
	HRP Conjugate Dilution Buffer
	Sample Diluent
	(20X) Wash Buffer
	TMB Substrate
	Stop Solution
	Consult Instructions for Use
	Catalog Number
	Expiration Date
	Storage Temperature